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Review

Genetic factors associated with the development of age-related macular degeneration

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ABSTRACT

Age-related macular degeneration (AMD) affects the macula and is the leading cause of significant and irreversible central visual loss. It is the most common cause of visual loss in people aged more than 60 years. This disease affects 2.5 million individuals in Europe. AMD is caused by both environmental and genetic factors. Numerous risk factors have been reported, but the pathogenesis of AMD is complex and fairly understood. Age, female gender, obesity, race, education status, family history, hyperopia, iris color, cigarette smoking, previous cataract surgery, history of cardiovascular and cerebrovascular disease, diabetes, sunlight exposure and many other factors have been shown to be associated with AMD development. Scientific evidence shows that genes may play a role in the development of nearly 3 out of 4 cases of this devastating eye disease. The genes that have been shown to be associated with AMD are genes encoding complement system components such as CFH, C2, C3, CFB, and other. © 2016 The Lithuanian University of Health Sciences. Production and hosting by Elsevier

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1. Introduction

Age-related macular degeneration (AMD) is referred to aging changes without any other obvious precipitating cause that occur in the central area of the retina (macula) in people aged 55 years and more [1]. Impairment of sight and blindness are debilitating and are among the three most feared medical conditions, after cancer and cardiovascular disease [2]. In developed countries, AMD is the most common cause of visual loss in persons aged more than 60 years [3]. More than 30% of adults >75 years-of-age have this disease; in \sim 6%–8% of these individuals, the disease progresses, causing the most severe degree of visual loss [4]. The prevalence of early AMD increased

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from 1.3% \pm 0.3% per subject in the 30-year-old to 40-year-old group, to 3.6% \pm 0.5% in the 41-year-old to 50-year-old group, to 7.9% \pm 0.9% in the 51-year-old to 60-year-old group, to 10.0% \pm 1.1% in the 61-year-old to 70-year-old group, to 8.3% \pm 0.2% in the 71-year-old to 80-year-old group, and to 8.0% \pm 5.5% in the >81-year-old group [5].

AMD is the third leading cause of blindness globally following cataract and glaucoma and it accounts for 8.7% of all blindness cases [6]. In Lithuania, AMD-related blindness accounted for 13.8% and took the second place in 2002 (Lithuanian Medical Social Expertise Commission). The number of AMD-affected people in the developed countries is increasing dramatically. In 2014, Wong et al. published a systematic literature review that aimed to estimate the number of people who will be affected by AMD in the future [7]. This study showed that the prevalence of any AMD was higher in Europeans than Asians and Africans (12.3% vs. 7.4% and 7.5%, respectively). Also, this study reported that the number of persons with AMD will globally increase to 196 million by 2020 and will reach 288 million by 2040 [7].

It is thought that AMD has a multifactorial etiology, the development of which may be caused by interrelation of environmental and body peculiarities; also genetic factors have an impact.

The aim of our article was to review literature, disclose the present view on the pathogenesis and classification of AMD, and reveal factors, especially genetic association, prognosis of the development of this disease.

2. Pathophysiology of age-related macular degeneration

Pathological changes in the macula, a special area of the retina, are associated with the development of AMD. The center of the macula is called the fovea and it contains a huge concentration of photoreceptors with cone cells, responsible for visual acuity and color perception, dominating (up to 200 000 cells/mm²) [8,9]. The parafoveal area is a region, where rode cells dominate, surrounding the fovea and permitting night vision. In the early stages of AMD, photoreceptors are mostly damaged in the parafovea [10].

Macular degenerative changes occur due to modification of the retinal pigment epithelium and drusen formation in the Bruch membrane, which consists of the retinal pigment epithelium and the choroidal choriocapillary layer. Drusen are extracellular small nodule-shape deposits, made of phospholipids, collagen, and neutral lipids [11]. Also, drusen contain zinc, carbohydrates, and at least 129 different proteins, including apolipoproteins (e.g., apoE, apoB) and excluding structural extracellular matrix. Approximately 30% of the drusen, with a core diameter of 15 µm, consist of nonesterified cholesterol, nonfibrillar amyloid, and peanut agglutininbinding carbohydrates [11]. These cores may represent nucleation sites for following deposition. Their accumulation alters the delivery of oxygen and nutrients, leading to lesions in the retinal pigment epithelium and a progressive degeneration of photoreceptors, while visual function impairment is associated with the amount of damaged photoreceptors [8].

Histologically drusen are divided into two types: soft and hard. Soft drusen are large, diffuse and composed of an amorphous, granular, and loose material with poorly defined borders [11]. Soft drusen, the main component of basal linear deposit is membranous debris, containing coiled membranes and vesicular outlines of putative retinal pigment epithelium origin. The deposit composition is solid neutral lipid-rich particles. These drusen can form exudative macular degeneration and later can induce the detachment of the neuroepithelium. If the process progresses, new vessel will grow, leading to exudative hemorrhage. Hard drusen are small, distinct, hemispherical, or round-shaped with well-defined borders. Druse composition is solid and hyalinized [11]. They have decreased RPE coverage, consistent with aberrant expression of amyloid A and vitronectin over small drusen. Usually, hard drusen consists of cholesterol, nonfibrillar amyloid, calcifications. Some of them may content nonesterified cholesterol-rich core, and some identified by more shells, which are rich in apoE, apoC-I, and esterified cholesterol [11]. Hard drusen can cause atrophy of the retinal pigment epithelium and the choriocapillary layer [8,9].

3. Classification of age-related macular degeneration

AMD is commonly categorized into early and late forms [8,11]:

- Early AMD is described by the presence of a large number of deposits known as drusen (<10), which appear below the retinal pigment epithelium and causing more or less pigmentation areas called hyperpigmentation or hypopigmentation. The pigmentation regions are generally diffused [8,11].
- Late form of AMD is classified into dry (with geographic atrophy of the retinal pigment epithelium with the lack of neovascularization areas) and wet types (or exudative; with new blood vessel formations in choroid, called the choroidal neovascularization areas, further leading to the formation or the disciform scars) [8]. The wet type is the heavier form of the disease than the other types [11]. It causes severe damage to the retina and more frequently leads to devastating consequences such as vision loss [8].

4. Risk factors

Epidemiological studies have shown a complex interplay among genetic predisposition, systemic factors, lifestyle and environmental risk factors associated with the risk of AMD development. Age is one of the highest and invariable factors; persons aged between 60 and 80 years are at a 3-fold greater risk of developing advanced AMD compared with those younger than 60 years [12]. Smoking is another significant and modifiable factor. A lot of studies have shown the influence of smoking on AMD formation and have demonstrated that previous and current smokers are inclined to develop AMD at least 5–10 years earlier than nonsmokers [13]. Other factors implicated in the development of AMD are gender, family predisposition, color of the iris, ethnicity, sunlight exposure, body mass index, eating habits, oxidative stress, inflammation and increased levels of inflammatory marker in blood, low antioxidant levels in blood and diet, cataract and its operation. All these factors considerably affect individual risk.

Age is the most significant risk factor for AMD development. Disease prevalence for late AMD can peak near 10% in persons older than 80 years [14]. The prevalence of late AMD is 1.4% (95% CI, 1.0%-2.0%) at the age of 70 years, rising to 5.6% (95% CI, 3.9%-7.7%) at age of 80 and 20% (95% CI, 14%-27%) at age of 90 [15]. Gender is the second important factor for AMD development: women are at potentially higher risk of neovascular AMD than men [15]. Family history is also a major risk factor for AMD [16,17]. The meta-analysis of the prospective cohort and crosssectional studies suggested that darker iris pigmentation (brown vs. blue eyes) was protective, but the overall results were not significant (OR = 0.88; 95% CI, 0.65-1.17 cross-sectional studies and RR = 0.98; 95% CI, 0.72-1.32 for prospective studies) [18]. Wong et al. found a higher prevalence of early and any AMD in Europeans than Asians (early: 11.2% vs. 6.8%, Bayes factor 3.9; any: 12.3% vs. 7.4%, Bayes factor 4.3), and early, late, and any agerelated macular degeneration to be more prevalent in Europeans than in Africans (early: 11.2% vs. 7.1%, Bayes factor 12.2; late: 0.5% vs. 0.3%, 3.7; any: 12.3% vs. 7.5%, 31.3) [7]. There was no difference in the prevalence between Asians and Africans (all Bayes factors <1 [19]. Europeans had a higher prevalence of geographic atrophy subtype (1.11%; 95% CI, 0.53%-2.08%) than Africans (0.14%, 0.04%-0.45%), Asians (0.21%, 0.04%-0.87%), and Hispanics (0.16%, 0.05%–0.46%) [7]. Among geographical regions, cases of early and any AMD were less prevalent in Asia than in Europe and North America (early: 6.3% vs. 14.3% and 12.8% [Bayes factor 2.3 and 7.6]; any: 6.9% vs. 18.3% and 14.3% [3.0 and 3.8]) [7]. Regarding sunlight as risk factors for AMD, controversial results have been reported: one study found no association between AMD and sun exposure or related factors except for the suggestion of an association between sunburn prone skin type and geographic atrophy, which reached borderline significance [19], while another study found that AMD was probably related to visible radiation especially blue light [20]. Cigarette smoking increases AMD risk. Current smoking was associated with an increased risk of transitioning from minimal to moderate early AMD [21]. Higher systolic blood pressure (OR = 1.06; 95% CI, 1.01-1.12 per 5 mmHg), overweight (OR = 2.87; 95% CI, 1.13-7.29), and obesity (OR = 2.92; 95% CI, 1.06-8.03), physical exercise duration (OR = 0.41; 95% CI, 0.18-0.96 for 30 min or more compared with less) and frequency (OR = 0.46; 95% CI, 0.23-0.92 for weekly or more often compared to less) were associated with late AMD in women only [22]. Also the prevalence of AMD is significantly higher in patients with myocardial infarction than in a random sample of the population [23]. Increased intake of fish reduced the risk of AMD, particularly for 2 or more servings per week (P trend = 0.04). Dietary omega-3 fatty intake was inversely associated with AMD (OR = 0.55; 95% CI, 0.32-0.95) comparing the highest vs. lowest quartile. Reduction in risk of AMD with higher intake of omega-3 fatty acids was seen primarily among subjects with low levels (below median) of linoleic acid intake, an omega-6 fatty acid (P trend < 0.001) [24]. Oxidative stress is believed to be a major mediator of the effect of age because mitochondrial oxidation is impaired with aging and oxidative damage is widely observed. Oxidative stress and the production of reactive oxygen species seem to play a pivotal role in AMD pathogenesis [25]. It is known that the macula receives the highest blood flow of any tissue in the body when related to size,

and anything that can reduce the rich blood supply can cause hypoxia, malfunction, or disease. Oxidative stress can affect both the lipid rich retinal outer segment structure and the light processing in the macula [25]. The levels of inflammatory markers, such as serum high-sensitivity C-reactive protein, tumor necrosis factor- α receptor 2, interleukin-6, and soluble vascular cell adhesion molecule-1, in blood were moderately related associated to the 20-year cumulative incidence of early AMD independent of age, smoking status, and other factors [26]. It is not known whether these associations represent a cause and effect relationship or whether other unknown confounders accounted for the findings. Even if inflammatory processes are a cause of early AMD, it is not known whether interventions that reduce systemic inflammatory processes will reduce the incidence of early AMD [26]. More recently, the data from the Age-Related Eye Disease Study concluded that cataract surgery was safe in the setting of dry AMD and did not accelerate progression to advanced sight threatening forms of AMD [27] and phacoemulsification surgery significantly improved vision in patients with neovascular AMD, with no increased need for anti-VEGF injections to keep the macula dry postoperatively [28]. Levels of serum vitamin D are inversely associated with early, but not advanced, AMD. Consistent use vs. non-use of vitamin D from supplements was inversely associated with early AMD only in individuals who did not consume milk daily [29]. Increased blood levels of homocysteine are associated with increased risk of AMD [30].

Not only environmental factors, but also genetic ones, have been reported to be associated with the development of AMD [31]. Over the past decade, more and more researchers focused their attention on detecting the genes, genetic components, which are concerned with the development of AMD. Familial studies have shown that family members of individuals with AMD are at a greater risk of developing the disease comparing to individuals having no family history of the disease [32]. Some twin studies have demonstrated pretty higher accordance rates between monozygotic twins as compared to accordance between dizygotic twins [33].

5. Genes associated with age-related macular degeneration

Genetic studies have shown that numerous genes may be related to age-related macular changes. Some strategies have been attempted to determine the exact genomic regions affected during AMD pathogenesis. The Retina International Database indicates 16 genes that have already been determined to have a link to AMD risk [34]. Complement factor H (CFH) was the first significant gene found to be related to AMD by a genome-wide association study. Complement factors B (CFB), 2 (C2), and 3 (C3) are other important genes highly associated with the risk of AMD development [35,36]. The second major locus of the risk of AMD development is linked to genes HTRA1 and ARMS2 [37]. In 2013, a genome-wide association study identified 7 new loci near genes COL8A1-FILIP1L, IER3-DDR1, SLC16A8, TGFBR1, RAD51B, ADAMTS9, and B3GALTL [38]. Eventually, our search combining all resources revealed 36 genes associated with AMD development (Table).

Table – Genes associated with the risk of age-related macular degeneration.							
Ge	ne	Name	Gene locus	Function	Variant/DNA marker	References	
1	ABCA1	ATP-binding cassette, sub- family A, member 1	1p22	Molecule transport across cell membranes	rs1883025	[39,40]	
2	ABCA4	ATP-binding cassette, sub- family A (ABC1), member 4	1p22		rs1800553, rs1800555	[41,42]	
3	ADAMTS9	ADAM metallopeptidase with	3p14.1	Regulation of angiogenesis suppression	rs6795735	[38]	
4	ARMS2	Age-related maculopathy	10q26	Unknown function, gene was found first in placenta and is located in the retina	rs10490924	[43]	
5	APOE	Apolipoprotein E	19q13.2	Lipid and cholesterol transport and catabolism	rs4420638	[38,42,44]	
6	B3GALTL	Beta 1,3-galactosyltransferase- like	13q12.3	Glycosylation pathway	rs9542236	[38]	
7	CETP	Cholesteryl ester transfer protein	16q21	Transport of cholesterol	rs3764261	[40,45]	
8	CFB	Complement factor B R32Q	6p21.3	Alternative complement activation	rs4151667, rs641153	[36,46]	
9	CFH	Complement factor H Y402H	1q32	Inhibitor of alternative complement	rs1061170	[46-48]	
		Independently associated single nucleotide polymorphism (SNP) variant	1q32	pairie,	rs10737680, rs1410996	[46,48]	
10	CFHR1	within intron 14 of CFH Complement factor H-related 1	1q31-q32	Possible overlapping function with CFH,	rs121913059	[49]	
11	CFHR3	Complement factor H-related 3	1q31-q32	complement regulation	84-kbp deletion	[49]	
12	CFI	Complement factor I	4q25	Complement cascade regulation	rs10033900, rs2285714	[50]	
13	COL8A1	Collagen type VIII, alpha1	3q12.3	Main component in basement membrane of the corneal endothelium	rs13095226	[39,45]	
14	COL10A1	Collagen, type X, alpha 1	6q21-q22	Produced by hypertrophic chondrocytes and located in mineralization zones of hyaline cartilage	rs3812111	[38]	
15	CX3CR1	Chemokine (C-X3-C motif) receptor 1	3p21	Leukocytes adhesive and migratory functions	rs3732378	[51,52]	
16	C2	Complement 2 E318D	9p21	Regulation of complement system activation	rs9332739	[36,46]	
17	C3 R1	Complement 3 R102G	19p13.3-p13.2	Regulation of complement system activation	rs2230199, rs1047286	[53–55]	
18	С9	Complement component 9	5p13.1	Regulator of the membrane Attack formation	rs34882957	[56]	
19	DDR1	Discodin domain receptor tyrosine kinase 1	6p21.3	Cell growth, differentiation and metabolism regulator	rs3094111, rs3130783	[38]	
20	ERCC6	Excision-repair cross- complementing, group 6	10q11	DNA transcription-coupled excision repair	rs3793784	[57]	
21	FBLN5	Fibulin 5	14q32.1	The extracellular matrix protein that promotes adhesion of endothelial cells	rs61734479	[58]	
22	FBLN6 or HMCN1	Fibulin-6 or hemicentin 1	1q24-q q31.1	Encodes a large extracellular member of the immunoglobulin superfamily	rs743137, rs680638	[59,60]	
23	FILIP1L	Filamin A interacting protein 1-like	3q12.1	Regulation of antiangiogenic activity in endothelial cells	rs13081855	[38]	
24	FRK	Fyn-related kinase	6q21-q22.3	Cell growth regulation	rs1999930, rs3812111	[38,61]	
25	HTRA1		10q26	Cell growth and insulin-like growth factors regulator	rs11200638	[62,63]	
26 27	IER3 LIPC	Immediate early response 3 Hepatic lipase C	6p21.3 15q22	Regulation of apoptosis Lipoprotein metabolism	rs3130783 rs10468017, rs493258, rs920915	[38] [45]	
28	QRX/RAXL1	Retina and anterior neural fold homeobox-like protein 1	19p13.3	Involved in development of the eye; possible modulation of the photoreceptor specific genes expression	Unknown	[64]	
29	RAD51B	RAD51 homolog B	14q23-q24.2	Pathway of DNA break repair	rs8017304	[38]	

Table (Continued)										
Gene		Name	Gene locus	Function	Variant/DNA References marker					
30	SLC16A8	Solute carrier family 16, member 8 (monocarboxylic acid transporter)	22q12.3-q13.2	Transport lactate across cell membrane	rs8135665	[38]				
31	TGFBR1	Transforming growth factor, beta receptor 1	9q22	Regulation of cell growth and division	rs334353	[38]				
32	TIMP3	TIMP metalloproteinase inhibitor 3	22q12.3	Extracellular matrix destruction	rs9621532, rs5749482	[40,45]				
33	TLR3	Toll-like receptor 3	4q35.1	Activation of innate immunity	rs3775291	[65–67]				
34	TLR4	Toll-like receptor 4	9q32-q33	Activation of innate immunity	rs4986790	[68]				
35	TNFRSF10A	Tumor necrosis factor receptor super-family, member 10a	8p21	Cell apoptosis regulator	rs13278062	[38]				
36	VEGFA	Vascular endothelial growth	6p12	Endothelial cell growth regulator;	rs943080,	[61]				
		factor A		angiogenesis	rs4711751,					
					rs833069,					
					rs1413711					

ABCA1. The study by Yu et al. examined 3066 individuals (of them, 221 were categorized as control subjects) and found a strong association between the ABCA1 gene and medium (P = 0.0044) and large (P = 0.00077) drusen as well as advanced AMD (P = 0.0003) [39].

ABCA4. Mutation in the ABCR gene was identified and had susceptibility in people with Stargardt disease. Two most common allele mutations ala1038 to val and gly1961 to glu were found [69]. In 1997, Allikmets et al. found first association of AMD at a significant level in people with AMD [70], and later in 1999, this was confirmed by the study of De La Paz et al. [71]. In 1999, Shroyer et al. performed mutation analysis in one family (family members of three generations) with AMD and Stargardt disease and identified heterozygous mutations in the ABCA4 gene. The authors proposed that some relatives of patients who carry Stargardt disease might have an increased risk of AMD development [72]. This was also confirmed by another study of the same authors in 2001 [73].

ARMS2. In 2000, Weeks et al. were the first to identify the link between the ARMS2 gene and susceptibility to AMD [74]. This gene is located on chromosome 10q26. Further research confirmed the association with AMD. In 2005, Rivera et al. identified the T allele of a single-nucleotide polymorphism (SNP) in the LOC387715 gene (rs10490924), which was related to the risk of AMD [75]. In 2006, the study by Schmidt et al. showed a statistically significant association between rs10490924 and a greater risk of AMD in smokers [76]. Kanda et al. reported that rs10490924 alone, the 200-kb region on chromosome 10q26, could account for susceptibility to AMD [77].

Fritsche et al. in 2013 confirmed the association between AMD and the T allele of rs10490924 with a P of 4×10^{-540} and OR of 2.76 [38].

The impact of MMP-2 Rs2285053 (C \rightarrow T), MMP-3 Rs3025039 (5A \rightarrow 6A), and MMP-9 Rs3918242 (C \rightarrow T) single nucleotide polymorphism on the development of early AMD was analyzed by the study of Liutkeviciene et al. It showed that only MMP-9 Rs3918242 (C \rightarrow T) single nucleotide polymorphism had a significant influence on the development of AMD, and it was more pronounced at the age of less than 65 years [78].

CFB. Gold et al. studied about 900 individuals with AMD and about 400 controls and reported that 26T-A transversion (resulting leu9 to his variant) and 95G-A transition (resulting arg32 to gln) in the CFB gene reduced the risk of AMD development [36]. The findings of the study by Maller et al. carried out in 2006 confirmed the association of leu9-to-his (L9H) and arg32-to-gln (R32Q) in the CFB gene with AMD [46].

CFH. Some genome studies on patients with AMD have revealed that polymorphisms in the CFH gene can be linked to the development of AMD [36,46]. CFH is a glycoprotein, circulating in blood plasma as an important hematologic component and regulating the activity of the complement system, protecting human cells and tissues from damage caused by pathogens and other dangerous substances during the stimulation of the system. The CFH gene is located on chromosome 1q32.

In 2005, Klein et al. performed genome-wide scans of 96 AMD cases and 50 controls to identify polymorphisms associated with AMD. Their analysis showed that an intronic and common variant was strongly related AMD ($P < 1 \times 10^{-7}$) and homozygous individuals harboring the risk allele were more than 7 times more likely to develop AMD (95% CI, 2.9-19) [47]. Another study by Edwards et al. investigated the locus for AMD (ARMD1) on chromosome 1q25-31 in 2 independent case-control cohorts: 224 cases and 134 controls. A significant association (P = 4.95×10^{-10}) was found within this locus, which has the gene encoding CFH, and the tyrosine-402 \rightarrow histidine-402 protein polymorphism was associated with a 2.7-fold greater risk of AMD [79]. In the study by Haines et al., Y402H, a common coding variant in the CFH gene, was found to increase the risk of AMD by 2.45-5.57 times [80]. In 2009, Bergeron-Sawitzke et al. examined 424 patients with AMD and 215 control individuals who were genotyped for single nucleotide polymorphisms. The study revealed that the GG genotype (rs1410996) of the CFH gene was associated with the greatest risk of AMD (OR = 6.6, 95% CI, 3.5–12; $P = 8.7 \times 10^{-11}$) [81].

In 2011, Ho et al. conducted a large-scale study including 2167 people genotyped for CFH Y402H or ARMS2A69S that aimed to determine whether the consumption of dietary nutrients can reduce the risk of early AMD. The study concluded that high intake of nutrients possessing antioxidant properties was associated with the reduced risk of early AMD [82]. Weismann et al. reported that the CFH polymorphism H402, known for its strong association with AMD, was capable to reduce the ability of the CFH gene to bind malondialdehyde, a lipid peroxidation product, that accumulates during numerous pathophysiological processes such as AMD [83]. The study by Hao et al. enrolled 109 AMD patients and 165 AMD-free controls and showed a significant link between CFH polymorphisms I62V and Y402H, and susceptibility to AMD in the Chinese population [84]. Carriers of the AA genotype and the A allele of I62V polymorphism were at 3.75-fold (95% CI, 1.70-8.30) and 1.64-fold (95% CI, 1.14-2.36) greater risk to develop AMD. For the Y402H polymorphism, the CT genotype and the C allele were associated with the higher risk of developing AMD (OR = 2.10, 95% CI, 1.04-4.27 and OR = 1.95, 95% CI, 1.02-3.72, respectively). Moreover, carriers of the AT haplotype were more likely than those with the GT haplotype to develop AMD (OR = 3.91, 95% CI = 2.58-5.94) [84].

CFHR1/CFHR3. CFHR1 and CFHR3 genes are located on chromosome 1q32 and regulate the activation of complement. The study by Hageman et al. carried out genotyping analyses of patients with AMD and controls and identified a large common deletion (copy number polymorphism 147 [CNP147]) that encompasses both these genes. The authors concluded that the absence of CFHR1 and/or CFHR3 might have a protective effect against AMD [85]. Another study showed that persons carrying fewer than 2 copies of CNP147 and haplotype T-G-D were at 1.75-fold (adjusted OR = 0.57; 95% CI, 0.38–0.8; P = 0.006) and 4-fold (OR = 0.25, 95% CI, 0.17–0.36; $P = 1.3 \times 10^{-13}$) lower risk of having AMD [86].

Raychaudhuri et al. matched the association of CFHR1/ CFHR3 deletion with AMD. They examined the Y402H allele, using rs10801555 as a proxy and rs1410996, using rs10737680 as a proxy. It was found that deletion of CFHR1/CFHR3 and rs10737680 decreased the risk of AMD, but they were not completely independent, assuming that there can be more variant, which were not yet recognized and can better explain the risk of disease [87].

CFI. The CFI gene encodes a complement system protein known as complement factor I. In 2013, van de Ven et al. analyzed and found 2 heterozygous missense mutations in the CFI gene: G188A and G119R. It was found the strong linkage to developing of AMD [88]. Especially, those patients who carried G119R mutation had more severe and advanced form of the disease. Also, 192 ancestry- and age-matched controls individuals, carrying the G188A and G119R mutation, did not have any other mutations in the CFH gene [88]. In addition, the G119R variant of CFI gene was recorded in persons with atypical hemolytic uremia syndrome [89]. Mild renal function decline in AMD-affected patients with G119R change of the CFI gene was noted. However, there was no significant difference in renal function between AMD individuals with G119R mutation and without it [90].

CX3CR1. In 2004, Tuo et al. examined 117 individuals with AMD and 276 controls and found that lower CX3CR1 expression was associated with AMD formation [91].

C2. The study involving nearly 900 individuals with AMD and nearly 400 controls showed that the variant of E318D and haplotype H7 in C2 considerably decreased the risk of developing AMD [36].

C3. It was reported that patients carrying K155Q variant (rs147859257) with the C allele of rs2230199 had an association

with high risk of AMD [38]. In 2013, Sedon et al. reported that the allele of the C3 gene encoding Gln155 (p.Lys155Gln substitution) caused resistance to proteolytic activation in the pathogenesis of AMD [92]. In the same year, the study by Helgason et al. confirmed this rare nonsynonymous SNP in the C3 gene. It was suggested that this substitution was associated with the reduction of C3 gene binding to CFH, resulting in resistance to inhibition by this factor that in turn leads to an increase in compliment activation [93].

C9. Recently, a new gene, increasing the risk of AMD, has been identified. In 2013, Seddon et al. performed genotyping of 5115 independent samples and confirmed an association between the allele in the C9 gene encoding p.Pro167Ser and AMD (joint OR = 2.2; $P = 6.5 \times 10^{-7}$) [92].

ERCC6. Tuo et al. examined a cohort of 460 individuals with advanced AMD and 265 controls and reported that C-6530 > G SNP in the ERCC6 gene was strongly related to AMD [57].

FBLN5. In 2004, Stone et al. examined 402 individuals with AMD and 429 controls and identified 7 various mutations in the FBLN5 gene in patients with confirmed AMD diagnosis and small circular drusen [58]. Another study examined 805 AMD-affected patients and 279 control cases and determined 9 FBLN5 gene mutations to be associated with different phenotypes of AMD and 2 associated with autosomal recessive cutis laxa. FBLN5 secretion was considerably decreased in cases of 4 AMD and 2 cutis laxa mutations. The findings of this study suggest that some missense mutations related to AMD result in reduced FBLN5 expression followed by a possible decrease in elastogenesis [94].

FBLN6. The study by Schultz et al. screened 20 candidate genes in the region between LAMB2 and D1S3469. Only one DNA variation in the hemicentin 1 (HMCN1) gene (16263A > G transition in exon 104 producing Gln5345Arg change) on chromosome 1q31 was identified in a large family with AMD [59]. Later, Fisher et al. reported that p.Gln5346Arg mutation, considered as a causal mutation, appeared to be documented at low frequency in the control group. These findings suggested that this polymorphism can provide the linkage to AMD in a small group of individuals, but did not lead essentially to the disease [95].

FRK/COL10A1. In 2011, Yu et al. performed a large metaanalysis of genome-wide association studies in order to identify genetic loci contributing to susceptibility to AMD. This study, besides confirmation of 10 loci previously reported to be associated with AMD, identified two novel susceptibility alleles, one of them being the rs1999930 allele near the FRK/ COL10A1 gene ($P = 1.1 \times 10^{-8}$) [61].

HTRA1. Dewan et al. reported that rs11200638 of the HTRA1 gene was the major genetic risk factor for the development of wet AMD and individuals carrying the risk genotype were 10 times more likely to develop this form of the disease than their counterparts with the wild-type genome [62]. Later, the HTRA1 gene (rs11200638) was confirmed to be associated with age-related macular degenerative disease ($P = 6.9 \times 10^{-29}$) [96].

LIPC. In 2013, Lee et al. analyzed 2 independent Caucasian cohorts: one including 1626 individuals with advanced AMD and 859 controls, and another with 2159 patients and 1150 control cases. rs493258 and rs10468017, two promoter variants of the *LIPC* gene, were found to have a link to advanced AMD in the Caucasian population [97].

QRX. Wang et al. investigated 92 cases with AMD; 322, with cone rod dystrophy; 14, with autosomal dominant retinitis pigmentosa; and 14, with autosomal recessive retinitis pigmentosa. There were 3 different heterozygous sequence variants in the QRX gene, one of which was found in a 67-yearold patient with AMD, who harbored a heterozygous G-to-A change (CGG to CAG, arginine to glutamine) at codon 87 [64].

TLR3. The study by Yang et al. analyzed associations between the TLR3 variant rs3775291 and advanced AMD (geographic atrophy and choroidal neovascularization) and reported that T allele (rs3775291) in the TLR3 gene had a protective effect against geographic atrophy, while none of the TLR3 variants was found to be associated with choroidal neovascularization [67].

VEGFA. Huang et al. performed a meta-analysis of 9 studies and showed that out of the 4 polymorphisms of the VEGFA gene, which were analyzed in this study, rs1413711 and rs833061 were found to be associated with an increased risk of AMD [98].

Other genes. In 2013, Fritsche et al. conducted a genomewide association study and for the first time identified 7 loci near the genes ADAMTS9, RAD51B, COL8A1/FILIP1L, IER3/DDR1, B3GALTL, SLC16A8, and TGFBR1. All these loci were associated with AMD at $P < 5 \times 10^{-8}$ [38]. The study by Whitmore et al. showed lower expression of the ADAMTS9 gene in samples high-risk CFH genotypes. ADAMTS9 has an influence on angiogenesis by suppressing it. Therefore, a decline in ADAMTS9 expression can be associated with a greater risk of developing AMD formation [99].

6. Concluding remarks

Numerous genes have been implicated in the pathogenesis of AMD. The genes associated with the complement system (CFH, CFB, CFI, C3) confer a higher risk of developing AMD. Moreover, understanding of genetic predisposition and the role of nongenetic environmental factors, such as diet, smoking, and other factors allows identifying biochemical processes for a greater part of AMD patients and essentially increases the opportunity to predict the risk of this disease and to develop new therapies. Despite current advanced technologies, the factors influencing the development of early and late AMD remain are still not entirely clear.

Conflict of interest

The authors state no conflict of interests.

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REFERENCES

[1] Ferris 3rd FL, Wilkinson CP, Bird A, Chakravarthy U, Chew E, Csaky K, et al. Clinical classification of age-related macular degeneration. Ophthalmology 2013;120(4):844–51.

- [2] Fritsche LG, Fariss RN, Stambolian S. Age-related macular degeneration: genetics and biology coming together. Annu Rev Genomics Hum Genet 2014;15:151–71.
- [3] Klein R, Peto T, Bird A, Vannewkirk MR. The epidemiology of age-related macular degeneration. Am J Ophthalmol 2004;137:486–95.
- [4] Friedman DS, O'Colmain BJ, Munoz B, Tomany SC, McCarty C, de Jong PT, et al. Prevalence of age-related macular degeneration in the United States. Arch Ophthalmol 2004;122:564–72.
- [5] Nangia V, Jonas JB, Kulkarni M, Matin A. Prevalence of agerelated macular degeneration in rural central India: the Central India Eye and Medical Study. Retina 2011;31: 1179–85.
- [6] Resnikoff S, Pascolini D, Etya'ale D, Kocur I, Pararajasegaram R, Pokharel GP, et al. Global data on visual impairment in the year 2002. Bull World Health Organ 2004;82:844–51.
- [7] Wong WL, Su X, Li X, Cheung CM, Klein R, Cheng CY, et al. Global prevalence of age-related macular degeneration and disease burden projection for 2020 and 2040: a systematic review and meta-analysis. Lancet Glob Health 2014;2(2): e106–16.
- [8] Joseph C, Bok BD. The retina and its disorders. San Diego, CA: Elsevier/Academic Press; 2011. p. 830–6.
- [9] Zarbin MA. Current concepts in the pathogenesis of agerelated macular degeneration. Arch Ophthalmol 2004;122:598–614.
- [10] Young RW. Pathophysiology of age-related macular degeneration. Surv Ophthalmol 1987;31:291–306.
- [11] Rudolf M, Clark ME, Chimento MF, Li CM, Medeiros NE, Curcio CA. Prevalence and morphology of druse types in the macula and periphery of eyes with age-related maculopathy. Invest Ophthalmol Vis Sci 2008;49:1200–9.
- [12] VanNewkirk M, Nanjan M, Wang J, Mitchell P, Taylor HR, McCarty CA. The prevalence of age-related maculopathy: the visual impairment project. Ophthalmology 2000;107 (8):1593–600.
- [13] Khan JC, Thurlby DA, Shahi H, Clayton DG, Yates JRW. Smoking and age related macular degeneration: the number of pack years of cigarette smoking is a major determinant of risk for both geographic atrophy and choroidal neovascularisation. Br J Ophthalmol 2006;90 (1):75–80.
- [14] Natl. Eye Inst.. Age-related macular degeneration (AMD). Bethesda, MD: Natl. Eye Inst.; 2014, http://www.nei.nih.gov/ eyedata/amd.asp.
- [15] Rudnicka AR, Jarrar Z, Wormald R, Cook DG, Fletcher A, Owen CG. Age and gender variations in age-related macular degeneration prevalence in populations of European ancestry: a meta-analysis. Ophthalmology 2012;119(3): 571–80.
- [16] Klein R, Cruickshanks KJ, Nash SD, Krantz EM, Nieto FJ, Huang GH, et al. The prevalence of age-related macular degeneration and associated risk factors. Arch Ophthalmol 2010;128:750–8.
- [17] Hyman L, Neborsky R. Risk factors for age-related macular degeneration: an update. Curr Opin Ophthalmol 2002;13:171–5.
- [18] Chakravarthy U, Wong TY, Fletcher A, Piault E, Evans C, Zlateva G, et al. Clinical risk factors for age-related macular degeneration: a systematic review and meta-analysis. BMC Ophthalmol 2010;10:31.
- [19] Khan JC, Shahid H, Thurlby DA, Bradley M, Clayton DG, Moore AT, et al. Age related macular degeneration and sun exposure, iris colour, and skin sensitivity to sunlight. Br J Ophthalmol 2006;90(1):29–32.
- [20] Yam JC, Kwok A. Ultraviolet light and ocular diseases. Int Ophthalmol 2014;34(2):383–400.

- [21] Myers CE, Klein BE, Gangnon R, Sivakumaran TA, Iyengar SK, Klein R. Cigarette smoking and the natural history of age-related macular degeneration: the Beaver Dam Eye Study. Ophthalmology 2014;121(10):1949–55.
- [22] Erke MG, Bertelsen G, Peto T, Sjølie AK, Lindekleiv H, Njølstad I. Cardiovascular risk factors associated with agerelated macular degeneration: the Tromsø Study. Acta Ophthalmol 2014;92(7):662–9.
- [23] Liutkeviciene R, Lesauskaite V, Zaliuniene D, Zaliaduonyte-Peksiene D, Cimbalas A, Jasinskas V, et al. Early age-related macular degeneration in patients with myocardial infarction. Curr Eye Res 2012;37(2):94–100.
- [24] Seddon JM, George S, Rosner B. Cigarette smoking, fish consumption, omega-3 fatty acid intake, and associations with age-related macular degeneration: the US Twin Study of Age-Related Macular Degeneration. Arch Ophthalmol 2006;124(7):995–1001.
- [25] Blasiak J, Petrovski G, Veréb Z, Facskó A, Kaarniranta K. Oxidative stress, hypoxia, and autophagy in the neovascular processes of age-related macular degeneration. Biomed Res Int 2014;2014:768026.
- [26] Klein R, Myers CE, Cruickshanks KJ, Gangnon RE, Danforth LG, Sivakumaran TA, et al. Markers of inflammation, oxidative stress, and endothelial dysfunction and the 20-year cumulative incidence of early age-related macular degeneration: the Beaver Dam Eye Study. JAMA Ophthalmol 2014;132(4):446–55.
- [27] Chew EY, Sperduto RD, Milton RC, Clemons TE, Gensler GR, Bressler SB, et al. Risk of advanced age-related macular degeneration following cataract surgery in the Age-Related Eye Disease Study: AREDS Report 25. Ophthalmology 2009;116(2):297–303.
- [28] Grixti A, Papavasileiou E, Balakrishna DC, Kumar V, Prasad S. Phacoemulsification surgery in eyes with neovascular age-related macular degeneration. ISRN Ophthalmol 2014;2014:417603.
- [29] Parekh N, Chappell RJ, Millen AE, Albert DM, Mares JA. Association between vitamin D and age-related macular degeneration in the Third National Health and Nutrition Examination Survey, 1988 through 1994. Arch Ophthalmol 2007;125:661–9.
- [30] Rochtchina E, Wang JJ, Flood VM, Mitchell P. Elevated serum homocysteine, low serum vitamin B12, folate, and agerelated macular degeneration: the Blue Mountains Eye Study. Am J Ophthalmol 2007;143:344–6.
- [31] Hyman LG, Lilienfeld AM, Ferris FL, Fine SL. Senile macular degeneration: a case–control study. Am J Epidemiol 1983;118(2):213–27.
- [32] Seddon JM, Ajani UA, Mitchell BD. Familial aggregation of age-related maculopathy. Am J Ophthalmol 1997;123: 199–206.
- [33] Grizzard SW, Arnett D, Haag SL. Twin study of age-related macular degeneration. Ophthalmic Epidemiol 2003;10: 315–22.
- [34] Retina international database: retinal and macular dystrophies; 2010 (Online). Available from: http://www. retina-international.org/sci-news/databases/ mutation-database/.
- [35] Francis PJ, Hamon SC, Ott J, Weleber RG, Klein ML. Polymorphisms in C2, CFB and C3 are associated with progression to advanced age related macular degeneration associated with visual loss. J Med Genet 2009;46:300–7.
- [36] Gold B, Merriam JE, Zernant J, Hancox LS, Taiber AJ, Gehrs K, et al. Variation in factor B (BF) and complement component 2 (C2) genes is associated with age-related macular degeneration. Nat Genet 2006;38:458–62.
- [37] Akagi-Kurashige Y, Yamashiro K, Gotoh N, Miyake M, Morooka S, Yoshikawa M, et al. MMP20 and ARMS2/HTRA1 are associated with neovascular lesion size in age-related

macular degeneration. Ophthalmology 2015;122(11). 2295–302.e2.

- [38] Fritsche LG, Chen W, Schu M, Yaspan BL, Yu Y, Thorleifsson G, et al. Seven new loci associated with agerelated macular degeneration. Nat Genet 2013;45:433–9.
- [39] Yu Y, Reynolds R, Fagerness J, Rosner B, Daly MJ, Seddon JM. Association of variants in the LIPC and ABCA1 genes with intermediate and large drusen and advanced age-related macular degeneration. Invest Ophthalmol Vis Sci 2011;52:4663–70.
- [40] Chen W, Stambolian D, Edwards AO. Genetic variants near TIMP3 and high-density lipoprotein-associated loci influence susceptibility to age-related macular degeneration. Proc Natl Acad Sci USA 2010;107:7401–6.
- [41] Allikmets R. Further evidence for an association of ABCR alleles with age-related macular degeneration. The International ABCR Screening Consortium. Am J Hum Genet 2000;67:487–91.
- [42] Katta S, Kaur I, Chakrabarti S. The molecular genetic basis of age-related macular degeneration: an overview. J Genet 2009;88:425–49.
- [43] Jakobsdottir J, Conley YP, Weeks DE, Mah TS, Ferrell RE, Gorin MB. Susceptibility genes for age-related maculopathy on chromosome 10q26. Am J Hum Genet 2005;77:389–407.
- [44] Klaver CC, Kliffen M, van Duijn CM, Hofman A, Cruts M, Grobbee DE, et al. Genetic association of apolipoprotein E with age-related macular degeneration. Am J Hum Genet 1998;63:200–6.
- [45] Neale BM, Fagerness J, Reynolds R, Sobrin L, Parker M, Raychaudhuri S, et al. Genome-wide association study of advanced age-related macular degeneration identifies a role of the hepatic lipase gene (LIPC). Proc Natl Acad Sci USA 2010;107:7395–400.
- [46] Maller J, George S, Purcell S, Fagerness J, Altshuler D, Daly MJ, et al. Common variation in three genes, including a noncoding variant in CFH, strongly influences risk of agerelated macular degeneration. Nat Genet 2006;38:1055–9.
- [47] Klein RJ, Zeiss C, Chew EY, Fagerness J, Altshuler D, Daly MJ, et al. Complement factor H polymorphism in age-related macular degeneration. Science 2005;308:385–9.
- [48] Rodríguez de Córdoba S, Esparza-Gordillo J, Goicoechea de Jorge E, Lopez-Trascasa M, Sánchez-Corral P. The human complement factor H: functional roles, genetic variations and disease associations. Mol Immunol 2004;41:355–67.
- [49] Raychaudhuri S, Iartchouk O, Chin K, Tan PL, Tai AK, Ripke S, et al. A rare penetrant mutation in CFH confers high risk of age-related macular degeneration. Nat Genet 2011;43:1232–6.
- [50] Fagerness JA, Maller JB, Neale BM, Reynolds RC, Daly MJ, Seddon JM. Variation near complement factor I is associated with risk of advanced AMD. Eur J Hum Genet 2009;17:100–4.
- [51] Combadiere C, Feumi C, Raoul W, Keller N, Rodéro M, Pézard A, et al. CX3CR1-dependent subretinal microglia cell accumulation is associated with cardinal features of age-related macular degeneration. J Clin Invest 2007;117:2920–8.
- [52] Ding X, Patel M, Chan CC. Molecular pathology of agerelated macular degeneration. Prog Retin Eye Res 2009;28: 1–18.
- [53] Maller JB, Fagerness JA, Reynolds RC, Neale BM, Daly MJ, Seddon JM. Variation in complement factor 3 is associated with risk of age-related macular degeneration. Nat Genet 2007;39:1200–1.
- [54] Yates JR, Sepp T, Matharu BK, Khan JC, Thurlby DA, Shahid H, et al. Complement C3 variant and the risk of agerelated macular degeneration. N Engl J Med 2007;357: 553–61.

- [55] Yang Z, Chen Y, Lillo C, Chien J, Yu Z, Michaelides M, et al. Mutant prominin 1 found in patients with macular degeneration disrupts photoreceptor disk morphogenesis in mice. J Clin Invest 2008;118:2908–16.
- [56] De Cordoba SR, de Jorge EG. Translational mini-review series on complement factor H: genetics and disease associations of human complement factor H. Clin Exp Immunol 2008;151(1):1–13.
- [57] Tuo J, Ning B, Bojanowski CM, Lin ZN, Ross RJ, Reed GF, et al. Synergic effect of polymorphisms in ERCC6 5' prime flanking region and complement factor H on age-related macular degeneration predisposition. Proc Natl Acad Sci USA 2006;103:9256–61.
- [58] Stone EM, Braun TA, Russell SR, Kuehn MH, Lotery AJ, Moore PA, et al. Missense variations in the fibulin 5 gene and age-related macular degeneration. New Eng J Med 2004;351:346–53.
- [59] Schultz DW, Klein M, Humpert AJ, Luzier CW, Persun V, Schain M, et al. Analysis of the ARMD1 locus: evidence that a mutation in HEMICENTIN-1 is associated with age-related macular degeneration in a large family. Hum Mol Genet 2003;12:3315–23.
- [60] Thompson CL, Klein BE, Klein R, Xu Z, Capriotti J, Joshi T, et al. Complement factor H and hemicentin-1 in age-related macular degeneration and renal phenotypes. Hum Mol Genet 2007;16:2135–48.
- [61] Yu Y, Bhangale TR, Fagerness J, Ripke S, Thorleifsson G, Tan PL, et al. Common variants near FRK/COL10A1 and VEGFA are associated with advanced age-related macular degeneration. Hum Mol Genet 2011;20:3699–709.
- [62] Dewan A, Liu M, Hartman S, Zhang SS, Liu DT, Zhao C, et al. HTRA1 promoter polymorphism in wet age-related macular degeneration. Science 2006;314:989–92.
- [63] Yang Z, Camp NJ, Sun H, Tong Z, Gibbs D, Cameron DJ, et al. A variant of the HTRA1 gene increases susceptibility to age-related macular degeneration. Science 2006;314: 992–3.
- [64] Wang Q, Chen S, Esumi N, Swain PK, Haines HS, Peng G, et al. QRX, a novel homeobox gene, modulates photoreceptor gene expression. Hum Mol Genet 2004;13:1025–40.
- [65] Allikmets R, Bergen AA, Dean M, Guymer RH, Hageman GS, Klaver CC, et al. Age-Related Macular Degeneration Genetics Consortium. Geographic atrophy in age-related macular degeneration and TLR3. N Engl J Med 2009;360:2252–4.
- [66] Edwards AO, Swaroop A, Seddon JM. Geographic atrophy in age-related macular degeneration and TLR3. N Engl J Med 2009;360:2254–5.
- [67] Yang Z, Stratton C, Francis P, Kleinman ME, Tan PL, Gibbs D, et al. Toll-like receptor 3 and geographic atrophy in agerelated macular degeneration. N Engl J Med 2008;359: 1456–63.
- [68] Zhang K, Kniazeva M, Han M, Li W, Yu Z, Yang Z, et al. A 5-bp deletion in ELOVL4 is associated with two related forms of autosomal dominant macular dystrophy. Nat Genet 2001;27:89–93.
- [69] Lewis RA, Shroyer NF, Singh N, Hosono N, Aoi T, Yasuda M, et al. Genotype/phenotype analysis of a photoreceptorspecific ATP-binding cassette transporter gene, ABCR, in Stargardt disease. Am J Hum Genet 1999;64:422–34.
- [70] Allikmets R, Shroyer NF, Singh N, Seddon JM, Lewis RA, Bernstein PS, et al. Mutation of the Stargardt disease gene (ABCR) in age-related macular degeneration. Science 1997;277(5333):1805–7.
- [71] De La Paz MA, Guy VK, Abou-Donia S, Heinis R, Bracken B, Vance JM, et al. Analysis of the Stargardt disease gene (ABCR) in age-related macular degeneration. Ophthalmology 1999;106:1531–6.

- [72] Shroyer NF, Lewis RA, Allikmets R, Singh N, Dean M, Leppert M, et al. The rod photoreceptor ATP-binding cassette transporter gene, ABCR, and retinal disease: from monogenic to multifactorial. Vis Res 1999;39:2537–44.
- [73] Shroyer NF, Lewis RA, Yatsenko AN, Wensel TG, Lupski JR. Cosegregation and functional analysis of mutant ABCR (ABCA4) alleles in families that manifest both Stargardt disease and age-related macular degeneration. Hum Mol Genet 2001;10:2671–8.
- [74] Weeks DE, Conley YP, Mah TS, Paul TO, Morse L, Ngo-Chang J, et al. A full genome scan for age-related maculopathy. Hum Mol Genet 2000;9:1329–49.
- [75] Rivera A, Fisher SA, Fritsche LG, Keilhauer CN, Lichtner P, Meitinger T, et al. Hypothetical LOC387715 is a second major susceptibility gene for age-related macular degeneration, contributing independently of complement factor H to disease risk. Hum Mol Genet 2005;14:3227–36.
- [76] Schmidt S, Hauser MA, Scott WK, Postel EA, Agarwal A, Gallins P, et al. Cigarette smoking strongly modifies the association of LOC387715 and age-related macular degeneration. Am J Hum Genet 2006;78:852–64.
- [77] Kanda A, Chen W, Othman M, Branham KE, Brooks M, Khanna R, et al. A variant of mitochondrial protein LOC387715/ARMS2, not HTRA1, is strongly associated with age-related macular degeneration. Proc Natl Acad Sci USA 2007;104:16227–32.
- [78] Liutkeviciene R, Lesauskaite V, Sinkunaite-Marsalkiene G, Zaliuniene D, Zaliaduonyte-Peksiene D, Mizariene V, et al. The role of matrix metalloproteinases polymorphisms in age-related macular degeneration. Ophthalmic Genet 2015;36(2):149–55.
- [79] Edwards AO, Ritter R, Abel KJ, Manning A, Panhuysen C, Farrer LA. Complement factor H polymorphism and agerelated macular degeneration. Science 2005;308:421–4.
- [80] Haines JL, Hauser MA, Schmidt S, Scott WK, Olson LM, Gallins P, et al. Complement factor H variant increases the risk of age-related macular degeneration. Science 2005;308:419–21.
- [81] Bergeron-Sawitzke J, Gold B, Olsh A, Lemon K, Visvanathan K, Allikmets R, et al. Multilocus analysis of age-related macular degeneration. Eur J Hum Genet 2009;17:1190–9.
- [82] Ho L, van Leeuwen R, Witteman JCM, van Duijn CM, Uitterlinden AG, Hofman A, et al. Reducing the genetic risk of age-related macular degeneration with dietary antioxidants, zinc, and omega-3 fatty acids: the Rotterdam Study. Arch Ophthalmol 2011;129:758–66.
- [83] Weismann D, Hartvigsen K, Lauer N, Bennett KL, Scholl HP, Charbel Issa P, et al. Complement factor H binds malondialdehyde epitopes and protects from oxidative stress. Nature 2011;478:76–81.
- [84] Hao XF, Xie LK, Tang YZ, Xie WK, Zhang ZF, Qi YX, et al. Association of complement factor H gene polymorphisms with age-related macular degeneration susceptibility. Exp Pathol 2015;8(3):3186–91.
- [85] Hageman GS, Hancox LS, Taiber AJ, Gehrs KM, Anderson DH, Johnson LV, et al. Extended haplotypes in the complement factor H (CFH) and CFH-related (CFHR) family of genes protect against age-related macular degeneration: characterization, ethnic distribution and evolutionary implications. Ann Med 2006;38:592–604.
- [86] Sawitzke J, Im KM, Kostiha B, Dean M, Gold B. Association assessment of copy number polymorphism and risk of agerelated macular degeneration. Ophthalmology 2011;118:2442–6.
- [87] Raychaudhuri S, Ripke S, Li M, Neale BM, Fagerness J, Reynolds R, et al. Associations of CFHR1–CFHR3 deletion and a CFH SNP to age-related macular degeneration are not independent. (Letter) Nat Genet 2010;42:553–5.

- [88] van de Ven JPH, Nilsson SC, Tan PL, Buitendijk GH, Ristau T, Mohlin FC, et al. A functional variant in the CFI gene confers a high risk of age-related macular degeneration. Nat Genet 2013;45:813–7.
- [89] Maga TK, Nishimura CJ, Weaver AE, Frees KL, Smith RJ. Mutations in alternative pathway complement proteins in American patients with atypical hemolytic uremic syndrome. Hum Mutat 2010;31:E1445–60.
- [90] Fakhouri F, Roumenina L, Provot F, Sallée M, Caillard S, Couzi L, et al. Pregnancy-associated hemolytic uremic syndrome revisited in the era of complement gene mutations. J Am Soc Nephrol 2010;21:859–67.
- [91] Tuo J, Smith BC, Bojanowski CM, Meleth AD, Gery I, Csaky KG, et al. The involvement of sequence variation and expression of CX3CR1 in the pathogenesis of age-related macular degeneration. FASEB J 2004;18(11):1297–9.
- [92] Seddon JM, Yu Y, Miller EC, Reynolds R, Tan PL, Gowrisankar S, et al. Rare variants in CFI, C3 and C9 are associated with high risk of advanced age-related macular degeneration. Nat Genet 2013;45:1366–70.
- [93] Helgason H, Sulem P, Duvvari MR, Luo H, Thorleifsson G, Stefansson H, et al. A rare nonsynonymous sequence variant in C3 is associated with high risk of age-related macular degeneration. Nat Genet 2013;45:1371–4.

- [94] Lotery AJ, Baas D, Ridley C, Jones RP, Klaver CC, Stone E, et al. Reduced secretion of fibulin 5 in age-related macular degeneration and cutis laxa. Hum Mutat 2006;27: 568–74.
- [95] Fisher SA, Rivera A, Fritsche LG, Keilhauer CN, Lichtner P, Meitinger T, et al. Case-control genetic association study of fibulin-6 (FBLN6 or HMCN1) variants in age-related macular degeneration (AMD). Hum Mutat 2007;28:406–13.
- [96] Fritsche LG, Loenhardt T, Janssen A, Fisher SA, Rivera A, Keilhauer CN, et al. Age-related macular degeneration is associated with an unstable ARMS2 (LOC387715) mRNA. Nat Genet 2008;40:892–6.
- [97] Lee J, Zeng J, Hughes G, Chen Y, Grob S, Zhao L, et al. Association of LIPC and advanced age-related macular degeneration. Eye (Lond) 2013;27(2):265–70.
- [98] Huang C, Xu Y, Li X, Wang W. Vascular endothelial growth factor A polymorphisms and age-related macular degeneration: a systematic review and meta-analysis. Mol Vis 2013;19:1211–21.
- [99] Whitmore SS, Braun TA, Skeie JM, Haas CM, Sohn EH, Stone EM, et al. Altered gene expression in dry age-related macular degeneration suggests early loss of choroidal endothelial cells. Mol Vis 2013;19:2274–97.